Towards relating the structure of Polyketide Synthases to their metabolic products

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Introduction

Polyketide-synthases (PKS) are a class of multi-domain megasynthases involved in the production of diverse of polyketides. The large class of PKS can be further divided into three different types (I-III) which differ in the way they biosynthesize its products. We have focused on type I PKS that either work iteratively (they use each domain several times) or in a multi-modular way. Modular PKS I are large enzymes composed of several modules, each containing a specific set of catalytic domains. They are similar to fatty-acid-synthases, but they can contain domains for both, total and partial reduction of its substrates.

However, relating the sequence of various catalytic domains present in a PKS biosynthetic gene cluster to the chemical structure of the final product is a challenging task [1,2].

In our studies we have addressed following questions: Can we predict the number of iterative steps catalyzed by an iterative PKS I? Is it possible to predict which substrate is accepted by a given acyl-transferase (AT) domain used for chain elongation?

Can we predict the number of elongation steps that are catalyzed by an iterative PKS I?

Methods

For prediction of the number of iterative steps being catalyzed by an iterative PKS I we measured the correlation between the cavity volume of the keto-synthase (KS) domain and the number of iterations. This methoad was previously proposed by Yadav et al [1]. Therefore homology models of different PKS I were built using the SBSPKS server. Each binding pocket was identified through superpositionig with 1B3N, a homologue β -ketoacyl-carrier-protein synthase that has been co-crystallized with an inhibitor. Cavity volume was measured using Sitemap (Schrödinger Inc.).



Fig. 1: Correlation between catalytic cavity volume and number of elongation steps for iterative and modular PKS I

Prediction of the substrate specificity of an AT domain Results Methods AT-domain with substrate specificity for methylmalonate affinity We calculated the three OŤ Predicting substrate specifity AT OT 3,2 representative three AT domains tor Comparison of the domains: docking

Results

Predicting the number of iterative condensation-steps:

A correlation was found between the cavity volume of the bindingpocket and the number of iterative steps being catalyzed by the corresponding KS domain (Fig.1). The correlation was weaker than previously reported. As expected no significant correlation was observed between the cavity volume of the last KS domain of modular PKS I and the size of its metabolic product.

candidate substrates (methylmalonate, malonate and methoxymalonate) using Glide 5.8 (Schrödinger Inc.). The AT domains were first modelled using Prime (Schrödinger Inc.). Templates were obtained from PDB after performing a BLAST search aiming at the identification of homologous proteins with available structural information. Afterwards, an all-atom energy minimization was carried out. To locate the binding pocket we performed a sequence motif search [2]. After docking with the three substrates we compared the corresponding docking scores to identify the favoured substrate. Furthermore, we performed a multiple sequence alignment using Jalview (http://www.jalview.org) to identify amino acids that are important for substrate specificity of AT domains.





AT-domain with substrate specificity for methoxymalonate





Fig. 2: Representation of the docking scores for each substrate and its specific AT domain and visualization of the binding mode of Methylmalonate in its corresponding binding pocket scores of different AT domains with possible substrates showed a slight correlation (Fig. 2) with the actual results. However it is to be considered that only elongation (Malonate, unit the Methylmalonate or Methoxymalonate) was used for docking so we cannot exclude that results would differ slightly if their CoA-derivatives were docked. Performing a multiple sequence alignment indicated that specific residues play a key role in substrate specificity for Methylmalonate or Malonate [2]. Moreover, we identified a sequence motif present in AT domains with specificity for Methoxymalonate (Fig. 3).

	10	70 80	90	100	210	220	230	240	250	260
Herbimycin1_methoxy/1-322	VEVEPGQGAQWAGM	TFAIMVSLAALWQAN	GIHPDÁVIGHS	SQGEIAAACVAGH	DFAGHSGHVDTIK	DQLHNVLDG	TATPGH TA	wmstvdadwa	NРТНІ ОР-	DYWYRNLRDTV
Herbimycin2_methoxy/1-322	VFVFPGQGAQWAGM	TFAIMVSLAALWQAN	GIHPDAVI <mark>G</mark> HS	SQGEIAAACVAGH	'DFAG <mark>H</mark> SGHVDTIK	DQLHNVLDG	TATPGH TA	WMSTVDADWA	NPTHIDP -	DYWYRNLRDTV
Tautomycin_methoxy/1-322	VFVFPGQGAQWAGM	SWAV <mark>MVSLAALW</mark> RSF	GVEPSAVV <mark>G</mark> H S	SQGELAAAVVGGY	'DFAGHSGHVDAIE	ERLRAELAD	TARPGE VP	WMSTVDGQWA	DHARVDA-	DYWYRNLRDVV
Concanamycin1_methoxy/1-320	VFVFPGQGS <mark>Q</mark> WVGM/	LFSMMVSLAALWRSY	GVEPSAVV <mark>G</mark> H S	SQGEIAAAVVAGA	TFAGHSPQVDEVR	GELLDALAG	APRRTD IA	FYSTVTGGVV	DTTTLDT -	- EYWYRNLREPV
Concanamycin2_methoxy/1-321	VFVFPGLGSQWPGM	LFSVMVSLAALWRSY	GVEPAAVV <mark>G</mark> H S	SQGEVAAACVAGA	'DYAS <mark>H</mark> SDDVSTVR	RDRLGEDLSSL	VPKAPAVP	LVSTVDADWI	GPGDLTH -	- EYWYRNLRQ TV
Soraphen2_methoxy/1-318	VFVFPGQGS <mark>Q</mark> WEGM/	LFSMMVSLAALWRSM	GVEPDAVV <mark>G</mark> H S	SQGEIAAACVAGA	DFASHSAQVESIR	RDELLDLLSWL	EPRSTA VP	FYSTVSGAAI	DGSELDA-	AYWYRNLRQPV
Soraphen1_methoxy/1-319	VFVFPGQGS <mark>Q</mark> WPGM#	LFTVMVSLAALWRSR	GIEPDAVV <mark>G</mark> HS	SQGELAAAYVAGA	'DVASHGAQIEGMR	REQLLEELREI	EPRESR IP	FYSTVRGEKL	AGTELGA -	AYWYDNLLRPV
HerbimycinA_methyl/1-320	VFVFPGQGAQWVGM	TFAVVVSLATLWQSM	GIHPDAVT <mark>G</mark> HS	SQGEIAAACVAGH	'DYASHTGHVDTIK	NELHQTLADI	TTEPGT LP	WLSTVDGEWI	EPDTLDS -	GYWYRNLRQTV
Chalcomycin_methyl/1-318	VFVFPGQGT <mark>Q</mark> WAGM/	SFAVMVSLAELWRSL	GVVPDAVV <mark>G</mark> H S	SQGEIAAAVVAGG	'DYASHSAHVEELR	RAELEQILAGI	DPVAGE TP	LYSTVEAGVV	DTASMDA-	GYWFRNLRRPV
Amphotericin_methyl/1-321	VFVFPGQGS <mark>Q</mark> WVGM(SFAVMVSLAAVWRAQ	GVEPDAVV <mark>G</mark> H S	SQGEIAAAVVSGA	'DYA <mark>SH</mark> SHHVEDLH	IDEILQLLAE	/APKASE VP	LFSTVTGDWL	DTTVMDA-	GYWFRSLRGRV
Chlorothricin_methyl/1-319	VFVFPGQGS <mark>Q</mark> WAGM/	LFAVMVSLAELWRSF	gvrpdavv <mark>g</mark> h §	SQGEIAAACVAGA	'DYAS <mark>H</mark> SHHVEAIR	RERLAELLAG	APRSCD VA	FYSTVYGEPV	DTGELDA -	GYWYRNLRDTV
ConcanamycinA_methyl/1-314	VFVFPGQGS <mark>Q</mark> WAGM/	LFAVMVSLAEVWRSF	GVVPDAVV <mark>G</mark> H S	SQGEIAAAVVAGA	'DYAS <mark>H</mark> SAHVEEIR	RETLLEALSGL	.RPTAAH VP	LYSTVEGGWL	DTARMDA -	- DYWYRNLRATV
Lasalocid_methyl/1-320	VFVFPGQGS <mark>Q</mark> WTGM(LFAVMVSLARLWQHH	GIHPDAVI <mark>G</mark> HS	SQGEIAAAHIAGA	'DYAS <mark>H</mark> SAQVESIR	RDTVLQAATGI	NPQPTT IP	LYSTVTGQPI	DGTQLDA -	- DYWYTN <mark>L</mark> RHTV
Monensin_methyl/1-316	VLVFPGQGS <mark>Q</mark> WVGM(LWAVMVSLAAVWADH	GVTPAAVV <mark>G</mark> H S	SQGEIAAVVVAGA	'DYAS <mark>H</mark> SPQVDAIT	DELTHTLSG	/RPTTAPVA	FYSAVTGTRI	DTAGLDT -	DYWVTNLRRPV
Nigericin_methyl/1-326	VLVFPGQGS <mark>Q</mark> WAGM(LWAAMVSLAAVWAEY	gvrppavv <mark>g</mark> h §	SQGEIAAAVVAGA	'NYAS <mark>H</mark> SPQVDEIA	HELIELLGG	/EPVEVSGSGVA	FYSTVTGGRA	DVSVLDT -	GYWVRNLRERV
Nanchangmycin_methyl/1-322	VMVFPGQGS <mark>Q</mark> WRGM(LWAVMVSLAAVWESY	GVTPTAVV <mark>G</mark> H S	SQGEIAAACVAGG	'DYA <mark>SH</mark> GPQVDRLA	DTIRTDLAD	.SPGASD AV	/FYSAVTGARQ	PTEELDA -	DYWF TNL RQ PV
Halstoctacosanolide_methyl/1-31	5 VFVFPGQGS <mark>Q</mark> WVGM(LWAVMVSLAAVWESW	GVVPAAVV <mark>G</mark> H S	SQGEIAAACVAGA	'DYA <mark>S</mark> HSAHMERIH	IDELLEILSG	EPKTSRIP	LYSTVSAARI	D T S R M D A -	SYWFDNIRGTV
Amphotericin4_malonyl/1-312	- F L F S G Q G S <mark>Q</mark> R L G M (LFAVEVALYRLVESW	GVKPDFVA <mark>G</mark> H S	SIGEIAAAHIAGV	SHAFHSPLMDPML	DEFRSVAEGL	.SYSAPAIP	VVSNLTGTLA	DPADLCSA	ADYWVRH MRDAV
Amphotericin5_malonyl/1-311	FSGQGS <mark>Q</mark> RLGM(LFAVEVALYRLVESW	GVRPDFVA <mark>G</mark> H S	SIGEIAAAHIAGV	SHAFHSPLMDLML	DEFRAVAETL	.SFAAPVIP	VVSNLTGSLA	TAEELCSP	PEYWVRHMREAV
Amphotericin3_malonyl/1-311	LFSGQGS <mark>Q</mark> RLGM(LFAVEVALYRLVSSL	GVTPDYVG <mark>G</mark> H S	SIGEIAAAHVAGV	SHAFHSPLMDPML	.EEFRRVARGL	. TYHEPR IP	VVSNLTGAIA	DPADLCTA	ADYWVRHMREAV
Herdamycin1_malonyl/1-313	LFPGQGA <mark>Q</mark> QTGA(LFALEVALFRLVESW	GIEPDVLI <mark>G</mark> HS	SVGELAAAHAAGI	SHAFHSRLMEPML	ARFAEVAEGL	.AYGAPRIP	VVSTLTGAVV	TDEAMSGA	SYWVRHARETV
Tautomycetin4_malonyl/1-268	LFTGQGA <mark>Q</mark> RVGM/	LFAVEAALFAVLRSY	GVRPAFLL <mark>G</mark> H S	SIGEVTAAYVAGV	SHAFHSALMDPML	.AEFARVLES\	/EFREPRIP	VVSNLTGVVG	DELTSP	GYWVRQMRGTV
Herdamycin0_malonyl/1-317	L F T G Q G G <mark>L</mark> R P G V (LFALETALYRLVCSL	GVRPALVA <mark>G</mark> H S	SVGEVAAAHAAGV	'THA <mark>FH</mark> SPLMEPVL	REFGRVCAGL	.SYRPPRVP	VVSTVTGRIA	AGTELCSF	PEYWVSHMRRPV
Oligomycin8_malonyl/1-320	LFPGQGA <mark>Q</mark> RPGM(LFALEVALFRLLASW	GVVPDYLL <mark>G</mark> H S	SVGE I AAAHAAGA	SHAFHSPLMEPML	DEFAELVAGL	.SFAPPRIP	VVSNLTGAVL	GADEFADF	RYWVRHARHTV
Oligomycin9_malonyl/1-331	- F L F T G Q G A Q R P G M (TFALGVALFRLLEEW	GVRPRLLS <mark>G</mark> H S	SVGELTAAHVSGM	SHAFHSPLMDPVV	/DPLRQVAARL	. T F G P P A I P	VVSSVTGTLL	EPAAWADF	PAYWARQ AREPV
Niddymycin1_malonyl/1-312	- L L F T G Q G A <mark>Q</mark> H R G M (LFALQTALYRTLTAR	G T Q A H L V L G H S	SVGEITAAHIAGV	SHAFHSALMDPML	GAFRDTLNTL	.NYQPPTIP	LISNLTGQIA	DPNHLCTP	PDYWIDHARHTV
Niddymycin3_malonyl/1-312	- L L F T G Q G A Q H P G M (LFALQTALYRTLTAR	G T Q A H L V L G H S	S <mark>VGE</mark> ITAAHIAGV	SHAFHSALMDPML	GAFRDTLNTL	.NYQPPTIP	LISNLTGQIA	DPNHLCTP	PDYWIDHARHTV

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Fig. 3: Residues shared by all AT-domains are marked in orange. Motifs present in AT-domains with substrate specificity for Methoxymalonate are enframed in blue. Residues that are specific for Methylmalonate are

highlighted in pink. Green frames indicate substrate specificity for Malonate.

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